

THE FLOWCHART BELOW ILLUSTRATES THE WORKFLOW TO BE FOLLOWED FOR BOTH URINE AND VAGINAL SWAB IN SPECIMEN TRANSPORT BUFFER



URINE

Take a 2ml sample

VAGINAL SWAB

Resuspend in 100µl RB

Nuclease-free
2ml microcentrifuge tube

1 SAMPLE CLEAN UP



Centrifuge
1 min @ 2700-3300 rcf



Discard Pellet Retain supernatant by pouring into a separate nuclease-free 2ml microcentrifuge tube

2 WASH



Centrifuge
5 min @ 2700-3300 rcf



Discard Supernatant Resuspend pellet in 1ml of CTNG Resuspension Buffer by adding buffer to the tube and pipetting up and down several times in the location of the pellet

3 RESUSPEND



Centrifuge
1 min @ 2700-3300 rcf



Discard Supernatant

URINE

Resuspend in 200µl RB

VAGINAL SWAB

Resuspend in 100µl RB

Note 1: During centrifugation steps, it may be useful to align the microcentrifuge tubes in the same direction, noted by the position of the tabs/hinges on the tube lids.

Note 2: Please refer to IFU page 11 for adjustment of sample volume for swab specimens.

4 LYSE

HEATBLOCK
10 mins @ 105oC

Remove and leave to equilibrate to RT for 5 mins

5 SPIN



Centrifuge
10 sec @2700-3300 rcf



Open the pouch containing the reaction strips.

If lyophilized pellets are sticking to the sealing film, tap the strip lightly until they fall to the bottom of the tubes.

Place the strip(s) in the HG Swift Set-up Rack in the correct orientation

6 RUN

Add 25µl of sample to Target and EC Wells

Close the lids firmly



Run HG Swift



- It is vital to orient the reaction tubes in the correct direction. When lids are facing away from the user, the left-hand tube in the strip contains Target reaction mix and the right-hand tube contains EC reaction mix.
- It is critically important to ensure that lids are fully closed before commencing the run.
- **Never** re-open the caps of the reaction strips after closing.
- After the run, remove the reaction tube strip from the HG Swift lifting by the handles of the Strip Carrier.
- Dispose of the used strips firstly into a small sealable plastic bag and then into a bin. Empty the bin regularly and do not allow large amounts of waste to build up on top of bags containing used reaction strips.
- Work areas must be regularly cleaned with appropriate DNA decontamination solutions.
- It is recommended to run NTCs periodically to check for contamination.

Note: If required, CT and NG can be added directly to reaction strips without heat treatment for positive control. Please check IFU for further details.